

IDKmonitor[®] Ustekinumab drug level ELISA

*For the in vitro determination of free ustekinumab
concentration (e. g. STELARA[®])
in EDTA plasma and serum*

Valid from 2021-07-12

REF KR9660

Σ
96

+8°C
+2°C

RUO



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1. INTENDED USE

The Immundiagnostik AG assay is an enzyme immunoassay intended for the quantitative determination of free therapeutic ustekinumab antibodies (e.g. STELARA®,) in EDTA plasma and serum. For *research* use only. Not for use in diagnostic procedures.

2. INTRODUCTION

Ustekinumab is a monoclonal, human therapeutic antibody targeting p40, the common subunit of interleukin 12 and 23 (IL-12/23). Ustekinumab blocks IL-12/23 signaling, thus inhibiting the t-cell mediated immune response [1].

The IDKmonitor® ELISA for the determination of the drug level of ustekinumab (e.g. STELARA®) measures quantitatively free ustekinumab.

3. MATERIAL SUPPLIED

Cat. No.	Label	Kit Components	Quantity
KR9660	PLATE	Microtiter plate, pre-coated	12 x 8 wells
KR0001.C.100	WASHBUF	Wash buffer concentrate, 10x	2 x 100 ml
KR9660	CONJ	Conjugate concentrate, peroxidase-labelled	1 x 200 µl
KR9660	STD	Standards, lyophilised (see specification for concentrations)	4 x 6 vials
KR9660	CTRL 1	Control, lyophilised (see specification for range)	4 x 1 vial
KR9660	CTRL 2	Control, lyophilised (see specification for range)	4 x 1 vial
KR9660	SAMPLEBUF	Sample dilution buffer, ready-to-use	2 x 100 ml
KR0002.15	SUB	Substrate (tetramethylbenzidin), ready-to-use	1 x 15 ml
KR0003.15	STOP	Stop solution, ready-to-use	1 x 15 ml

For reorders of single components, use the catalogue number followed by the label as product number.

4. MATERIAL REQUIRED BUT NOT SUPPLIED

- Ultrapure water*
- Calibrated precision pipettors and 10–1000 µl single-use tips
- Foil to cover the microtiter plate
- Horizontal microtiter plate shaker
- Multi-channel pipets or repeater pipets
- Centrifuge
- Vortex
- Standard single-use laboratory glass or plastic vials, cups, etc.
- Microtiter plate reader (required filters see chapter 7)

* Immundiagnostik AG recommends the use of ultrapure water (water type 1; ISO 3696), which is free of undissolved and colloidal ions and organic molecules (free of particles > 0.2 µm) with an electrical conductivity of 0.055 µS/cm at 25 °C (≥ 18.2 MΩ cm).

5. STORAGE AND PREPARATION OF REAGENTS

- To run the assay more than once, ensure that reagents are stored at the conditions stated on the label. **Prepare only the appropriate amount necessary for each run.** The kit can be used up to 4 times within the expiry date stated on the label.
- Reagents with a volume less than **100 µl** should be centrifuged before use to avoid loss of volume.
- **Preparation of the wash buffer:** The **wash buffer concentrate (WASHBUF)** has to be diluted with ultrapure water **1:10** before use (100 ml WASHBUF + 900 ml ultrapure water), mix well. Crystals could occur due to high salt concentration in the concentrate. Before dilution, the crystals have to be redissolved at room temperature or in a water bath at 37 °C. The **WASHBUF** can be used until the expiry date stated on the label when stored at **2–8 °C**. **Wash buffer** (1:10 diluted WASHBUF) can be stored in a closed flask at **2–8 °C for 1 month**.
- The **lyophilised standards (STD)** and **controls (CTRL)** can be used until the expiry date stated on the label when stored at **2–8 °C**. **Reconstitution** details are given in the **specification data sheet**. **Standards and controls** (reconstituted STD and CTRL) **are not stable and cannot be stored**.
- **Preparation of the conjugate:** Before use, the **conjugate concentrate (CONJ)** has to be diluted **1:101** in wash buffer (100 µl CONJ + 10 ml wash buffer). can be used until the expiry date stated on the label when stored at **2–8 °C**. **Conjugate** (1:101 diluted CONJ) **is not stable and cannot be stored**.

- All other test reagents are ready-to-use. Test reagents can be used until the expiry date (see label) when stored at **2–8 °C**.

6. STORAGE AND PREPARATION OF SAMPLES

Sample storage

Freshly collected EDTA plasma or serum can be stored for 7 days at room temperature (22–23 °C) or 4 °C and for up to 6 months at -20 °C.

Sample preparation

EDTA plasma or serum samples must be diluted **1:200** before performing the assay, e.g.

10 µl sample + **1 990 µl** sample dilution buffer (SAMPLEBUF), mix well.

For testing in duplicates, pipet **2 x 100 µl** per well of each prepared sample.

7. ASSAY PROCEDURE

Principle of the test

This ELISA is designed to determine the quantity of free ustekinumab (therapeutic antibody against interleukins 12 and 23) in EDTA plasma or serum samples. In a first incubation step, the free ustekinumab from the sample is bound to the specific monoclonal anti-ustekinumab antibody coated on the plate. After a washing step, ustekinumab bound to the plate is detected by adding a peroxidase-labelled anti-ustekinumab antibody. Tetramethylbenzidine (TMB) is used as a substrate for peroxidase. Finally, an acidic stop solution is added to terminate the reaction. The colour changes from blue to yellow. The intensity of the yellow colour is directly proportional to the concentration of free ustekinumab in the sample. A dose response curve of the absorbance unit (optical density, OD) vs. concentration is generated, using the values obtained from standard. The concentrations of free ustekinumab in the samples are determined directly from this curve.

Test procedure

Bring all **reagents and samples to room temperature** (15–30 °C) and mix well.

Take as many microtiter strips as needed from the kit. Store unused strips together with the desiccant bag in the closed aluminium packaging at 2–8 °C. Strips are stable until expiry date stated on the label.

For automated ELISA processors the given protocol may need to be adjusted according to the specific features of the respective automated platform. For further details please contact your supplier or Immundiagnostik AG.

We recommend to carry out the tests in duplicate.

1.	Add each 100 µl standards/controls/diluted samples into the respective wells.
2.	Cover the strips and incubate for 1 hour at room temperature (15–30°C) on a horizontal shaker* .
3.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
4.	Add 100 µl conjugate (diluted CONJ) into each well.
5.	Cover the strips and incubate for 1 hour at room temperature (15–30°C) on a horizontal shaker* .
6.	Discard the content of each well and wash 5 times with 250 µl wash buffer . After the final washing step, remove residual wash buffer by firmly tapping the plate on absorbent paper.
7.	Add 100 µl substrate (SUB) into each well.
8.	Incubate for 10–20 min** at room temperature (15–30°C) in the dark .
9.	Add 100 µl stop solution (STOP) into each well and mix well.
10.	Determine absorption immediately with an ELISA reader at 450 nm against 620 nm (or 690 nm) as a reference. If no reference wavelength is available, read only at 450 nm. If the extinction of the highest standard exceeds the range of the photometer, absorption must be measured immediately at 405 nm against 620 nm as a reference.

* We recommend shaking the strips at 550 rpm with an orbit of 2 mm.

** The intensity of the colour change is temperature sensitive. We recommend observing the colour change and stopping the reaction upon good differentiation.

8. RESULTS

The following algorithms can be used alternatively to calculate the results. We recommend using the 4 parameter algorithm.

1. 4 parameter algorithm

It is recommended to use a linear ordinate for the optical density and a logarithmic abscissa for the concentration. When using a logarithmic abscissa, the zero standard must be specified with a value less than 1 (e.g. 0.001).

2. Point-to-point calculation

We recommend a linear ordinate for the optical density and a linear abscissa for the concentration.

3. Spline algorithm

We recommend a linear ordinate for the optical density and a linear abscissa for the concentration.

The plausibility of the duplicate values should be examined before the automatic evaluation of the results. If this option is not available with the programme used, the duplicate values should be evaluated manually.

EDTA-plasma and serum samples

The obtained results have to be multiplied by the **dilution factor of 200** to get the actual concentrations.

In case **another dilution factor** has been used, multiply the obtained result by the dilution factor used.

9. LIMITATIONS

Samples with concentrations above the measurement range (see definition below) can be further diluted and re-assayed. Please consider this higher dilution when calculating the results.

Samples with concentrations lower than the measurement range (see definition below) cannot be clearly quantified.

The upper limit of the measurement range can be calculated as:

highest concentration of the calibration curve × sample dilution factor to be used

The lower limit of the measurement range can be calculated as:

LoQ × sample dilution factor to be used

LoQ see chapter "Performance Characteristics".

10. QUALITY CONTROL

Immundiagnostik AG recommends the use of external controls for internal quality control, if possible.

Control samples should be analysed with each run. Results, generated from the analysis of control samples, should be evaluated for acceptability using appropriate statistical methods. The results for the samples may not be valid if within the same assay one or more values of the quality control sample are outside the acceptable limits.

Reference range

We recommend each laboratory to establish its own reference range.

11. PERFORMANCE CHARACTERISTICS

Accuracy – Precision

Repeatability (Intra-Assay); n = 20

The repeatability was assessed with 3 serum samples under **constant** parameters (same operator, instrument, day and kit lot).

Sample	Mean value [ng/ml]	CV [%]
1	116.30	5.7
2	54.94	3.0
3	25.52	2.6

Reproducibility (Inter-Assay); n = 52

The reproducibility was assessed with 3 serum samples under **varying** parameters (different operators, instruments, days and kit lots).

Sample	Mean value [ng/ml]	CV [%]
1	113.23	6.3
2	54.95	5.3
3	27.15	5.0

Analytical sensitivity

The following values have been estimated based on the concentrations of the standard curve without considering possibly used sample dilution factors.

Limit of blank, LoB

0.503 ng/ml

Limit of quantitation, LoQ

1.470 ng/ml

Linearity

The linearity states the ability of a method to provide results proportional to the concentration of analyte in the test sample within a given range. This was assessed according to CLSI guideline EP06-A with a serial dilution of 2 different serum samples.

For Ustekinumab in serum and plasma, the method has been demonstrated to be linear from 4.35 to 299.00 ng/ml, showing a non-linear behaviour of less than $\pm 20\%$ in this interval.

Sample	Dilution	Expected [ng/ml]	Obtained [ng/ml]	Recovery [%]
A	1:200	300.00	299.00	100
	1:400	150.00	151.05	101
	1:800	75.00	71.10	95
	1:1 600	37.50	32.80	87
	1:3 200	18.75	16.75	89
	1:6 400	9.37	9.20	98
	1:12 800	4.68	5.05	108
B	1:200	300.00	295.35	98
	1:400	150.00	143.85	96
	1:800	75.00	70.00	93
	1:1 600	37.50	31.40	84
	1:3 200	18.75	16.10	86
	1:6 400	9.37	8.35	89
	1:12 800	4.68	4.35	93

Analytical specificity

The specificity of the antibody was tested by measuring the cross-reactivity against a range of compounds with structural similarity to Ustekinumab. There was no cross-reactivity observed.

Substance tested	Concentration added [ng/ml]	Concentration obtained [ng/ml]	Conclusion
Infliximab	4.2–225	< 0.503	< LoB
Adalimumab	4.2–225	< 0.503	< LoB
Golimumab	4.2–225	< 0.503	< LoB
Etanercept	4.2–225	< 0.503	< LoB
Vedolizumab	15.6–1000	< 0.503	< LoB

12. PRECAUTIONS

- All reagents in the kit package are for *research* use only.
- Kit reagents contain sodium azide or ProClin as bactericides. Sodium azide and ProClin are toxic. Substrates for the enzymatic colour reactions are toxic and carcinogenic. Avoid contact with skin or mucous membranes.
- The 10x Wash buffer concentrate (WASHBUF) contains surfactants which may cause severe eye irritation in case of eye contact



Warning: Causes serious eye irritation

IF IN EYES: Rinse cautiously with water for several minutes. Remove contact lenses, if present and easy to do. Continue rinsing. If eye irritation persists: get medical Advice/attention.

- The stop solution consists of diluted sulphuric acid, a strong acid. Although diluted, it still must be handled with care. It can cause burns and should be handled with gloves, eye protection, and appropriate protective clothing. Any spill should be wiped up immediately with copious quantities of water. Do not breath vapour and avoid inhalation.

13. TECHNICAL HINTS

- Do not interchange different lot numbers of any kit component within the same assay. Furthermore we recommend not assembling wells of different microtiter plates for analysis, even if they are of the same batch.
- Control samples should be analysed with each run.
- Reagents should not be used beyond the expiration date stated on the kit label.
- Substrate solution should remain colourless until use.
- To ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary.
- Avoid foaming when mixing reagents.
- Do not mix plugs and caps from different reagents.
- The assay should always be performed according to the enclosed manual.












14. GENERAL NOTES ON THE TEST AND TEST PROCEDURE

- The guidelines for laboratories should be followed.
- *IDKmonitor*® is a trademark of Immundiagnostik AG.
- Incubation time, incubation temperature and pipetting volumes of the components are defined by the producer. Any variation of the test procedure, which is not coordinated with the producer, may influence the results of the test. Immundiagnostik AG can therefore not be held responsible for any damage resulting from incorrect use.
- Warranty claims and complaints regarding deficiencies must be logged within 14 days after receipt of the product. The product should be sent to Immundiagnostik AG along with a written complaint.

15. REFERENCES

1. Scherl EJ, Kumar S & Warren RU (2010) Review of the safety and efficacy of ustekinumab. *Therap. Adv. Gastroenterol.* **3**: 321–328.
2. Chiu HY, Chu TW, Cheng YP & Tsai TF (2015) The association between clinical response to ustekinumab and immunogenicity to ustekinumab and prior adalimumab. *PLoS One* **10**: 1–10.

Used symbols:

	Temperature limitation		Catalogue number
	For research use only		To be used with
	Manufacturer		Contains sufficient for <n> tests
	Lot number		Use by
	Attention		Consult instructions for use
	Consult specification data sheet		

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